

Accuracy of procalcitonin for sepsis diagnosis in critically ill patients: systematic review and meta-analysis

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Procalcitonin is widely reported as a useful biochemical marker to differentiate sepsis from other non-infectious causes of systemic inflammatory response syndrome. In this systematic review, we estimated the diagnostic accuracy of procalcitonin in sepsis diagnosis in critically ill patients. 18 studies were included in the review. Overall, the diagnostic performance of procalcitonin was low, with mean values of both sensitivity and specificity being 71% (95% CI 67–76) and an area under the summary receiver operator characteristic curve of 0.78 (95% CI 0.73–0.83). Studies were grouped into phase 2 studies (n=14) and phase 3 studies (n=4) by use of Sackett and Haynes' classification. Phase 2 studies had a low pooled diagnostic odds ratio of 7.79 (95% CI 5.86–10.35). Phase 3 studies showed significant heterogeneity because of variability in sample size (meta-regression coefficient -0.592 , $p=0.017$), with diagnostic performance upwardly biased in smaller studies, but moving towards a null effect in larger studies. Procalcitonin cannot reliably differentiate sepsis from other non-infectious causes of systemic inflammatory response syndrome in critically ill adult patients. The findings from this study do not lend support to the widespread use of the procalcitonin test in critical care settings.

Introduction

Sepsis is the leading cause of mortality in critically ill patients.¹ Delay in diagnosis and treatment often results in rapid progression to circulatory collapse, multiple organ failure, and eventually death.^{2,3} Therefore, accurate and timely diagnosis will limit morbidity, reduce costs, and improve patients' outcome.^{4–6}

The diagnosis of sepsis is difficult, because clinical signs of sepsis often overlap with other non-infectious causes of systemic inflammation.^{7,8} These signs include tachycardia, leucocytosis, tachypnoea, and pyrexia, which are collectively termed a systemic inflammatory response syndrome (SIRS). SIRS is very common in critically ill patients, being found in various conditions including trauma, surgery, and hypoxic injuries.^{8–11} Microbiological culture can be used to distinguish sepsis from non-infectious conditions. However, this method lacks sensitivity and specificity, and there is often a substantial time delay.¹²

Procalcitonin, a 116-aminoacid peptide involved as a precursor in calcium homeostasis, has been studied as a marker to differentiate sepsis from other non-infectious causes of SIRS. Early studies were encouraging,^{13–16} and procalcitonin has been proposed as a diagnostic marker to be included in the international definition of sepsis.¹⁷ However, more recent studies have produced conflicting results.^{18–24} Furthermore, many studies included patients who did not have SIRS or who were not critically ill. This has added further uncertainty in assessing the diagnostic accuracy of procalcitonin in the critical care setting. The aim of this review was therefore to systematically and quantitatively evaluate all the published studies that assessed the diagnostic use of procalcitonin in critical care settings.

Methods

Data source

We searched Medline, Embase, and Current Contents from January, 1966, to November, 2005, for all studies of

diagnostic accuracy of procalcitonin for sepsis. The search strategy used medical subject heading terms and text words, including the following: “procalcitonin”; “sepsis”, “sepsis syndrome”, “septicemia”, “infection”, “systemic inflammatory response syndrome”, and “SIRS”; and “sensitivity”, “specificity”, “predictive value”, “likelihood ratio”, “review”, “meta-analysis”, “false positive”, and “false negative”.

The reference lists of each primary study were searched for additional publications. Further searches were done by manually reviewing abstract booklets, conference proceedings, and review articles. Investigators were contacted for further study details if needed. No language restriction was used and all foreign language publications were translated.

Study eligibility

We included all studies that met the following criteria: assessed the diagnostic accuracy of procalcitonin for sepsis; provided sufficient information to construct the 2x2 contingency table; and had a well-defined reference standard for the target condition (sepsis), which included use of accepted definitions by the American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference,⁸ and confirmed the presence of infection by microbiological culture.

Studies were excluded if they included patients who did not have SIRS or were not critically ill; included too narrow spectrum of patients, such as abdominal sepsis or septic shock; were duplicated studies; were paediatric studies; were limited to very restrictive subgroups, such as cardiac surgery, pancreatitis, meningitis, or burns; or were risk stratification or prognosis studies.

Data extraction

Two reviewers (BMPT, GDE) independently abstracted data in each study to obtain information on year of publication, country of origin, clinical setting, patients'

Methodological variable	Information required in each study	Studies that met criteria (n)
Did investigators use additional information (other than consensus definition of sepsis and microbiological culture) to confirm diagnosis, thus minimising misclassification bias?	Using all available information to diagnose sepsis/SIRS, including images studies, response to antibiotics, necropsy reports, and surgical findings	14
Was there a time delay between the index test and reference test (disease progression bias)?	Both procalcitonin and reference test to be done at the same time	18
Did the result of index test influence whether patients receive reference test (work-up bias)?	All patients should receive reference test regardless of procalcitonin test results	18
Were different reference tests used in patients (differential verification bias)?	Consistent use of international consensus criteria to diagnose sepsis in all patients	18
Was the interpretation of the reference test made without the knowledge of the index test (blinding)?	Diagnosis of sepsis/SIRS was made independent of the result of procalcitonin test	8
Description of reference test	Sufficient details provided in how the diagnosis was made	18
Description of index test	Sufficient details provided in how the procalcitonin was measured	18
Description of study population	Sufficient details provided for the case mix and demographic information of the patients enrolled	18
Method of recruitment	Patients were prospectively or consecutively recruited	18

SIRS=systemic inflammatory response syndrome.

Table 1: Quality assessment of the 18 studies included, by methodological variable

demographics, sample size, diagnostic cut-off points, and disease prevalence. Each reviewer extracted the data to construct a 2x2 table. Any disagreements were resolved by consensus.

Quality assessment

The methodological quality of each study was assessed by a checklist, by use of adapted criteria from the Cochrane Collaboration guidelines,²⁵ a study by Lijmer and colleagues,²⁶ and the QUADAS tool.^{27,28} Details of the methodological assessment are shown in table 1.

Statistical analysis

Studies were grouped according to Sackett and Haynes' classification of diagnostic studies.²⁹ In this classification, phase 1 studies are those that compare the difference in test results between patients with the target disorder and healthy individuals. Phase 2 studies are those that examine how the index test discriminates between patients with and without the target disorder. Phase 3 studies are those that assess the test's real-life performance in patients suspected to have the disorder.

For each study, positive and negative likelihood ratios and a diagnostic odds ratio (OR) were calculated. The likelihood ratio expresses the magnitude by which the probability of sepsis in a given patient is modified by the results of the procalcitonin test. It incorporates both sensitivity and specificity and has the advantage of being less affected by prevalence. The diagnostic OR is the ratio of the odds of a positive result in a patient with sepsis compared with a patient without sepsis: $[\text{sensitivity}/(1-\text{sensitivity})]/[(1-\text{specificity})/\text{specificity}]$. The diagnostic OR is a measure of overall accuracy and has the advantage of allowing the inclusion of covariates to examine heterogeneity in a regression model.³⁰ Pooling of the

summary indices was done using DerSimonian and Laird's random-effects model.³¹ Each study was weighted by use of an inverse variance method.

To detect heterogeneity, the likelihood ratios and diagnostic ORs were graphically displayed using forest plots and analysed using Cochran's *Q* test. A *p* value of less than 0.05 by Cochran's *Q* test indicated significant heterogeneity. To quantify the extent of heterogeneity, the *I*² statistic was used to measure the percentage of variability among summary indices that were caused by heterogeneity rather than chance. A study with an *I*² greater than 50% indicated substantial heterogeneity.

We constructed summary receiver operator characteristic (SROC) curves to summarise the study results, by use of a regression model described by Littenberg and Moses.³² In this method, the true-positive and false-positive rates of each study were logarithmically transformed and calculated in a regression model. The data were then back-transformed into the SROC space. A smoothed curve was then fitted across studies to represent the relation between sensitivity and the proportion of false positives (1-specificity).

To ensure that variation in the diagnostic threshold did not affect the shape of the SROC curve, the threshold effect was tested using the regression equation $D=a+bS$, where *D* is the log of the diagnostic OR and *S* is a measure of the diagnostic threshold. Estimation of the variables *a* and *b* was then done using a least-squares method, weighted by inverse variance. The absence of a threshold effect was indicated by $b=0$.

A *Q** point on the SROC curve was used to obtain the maximum joint sensitivity and specificity. The *Q** point is the intersection between a symmetrical SROC curve and the antidiagonal line, at which sensitivity equals specificity. This point represents a single-number summary of the

test performance and has the advantage of being less affected than other parameters by heterogeneity.^{32,33}

To explore sources of heterogeneity among studies, the Littenberg-Moses method³² was extended by adding covariates to the model. The covariates included spectrum characteristics (eg, study setting, prevalence), clinical and demographic variables (eg, disease severity, age), and methodological features (eg, sample size).

Publication bias was examined visually by inspecting funnel plots and statistically by using Egger's regression model.³⁴ If publication bias was present, the effect of such bias on the final summary estimate was assessed by using the trim and fill method.³⁵ This method imputes the missing studies and re-calculates a new summary estimate. The difference between the calculated and observed value was then used to determine the effect of bias on the diagnostic performance of the test.

Results

Study characteristics

We retrieved 672 abstracts, of which 39 were considered potentially suitable. After full text review, 21 studies were excluded (figure 1): one had no SIRS patients in the control group,³⁶ four included patients who were not critically ill,³⁷⁻⁴⁰ two were case-control studies,^{41,42} three used a different reference standard,⁴³⁻⁴⁵ nine could not generate 2x2 tables,^{14,46-53} and two had too narrow a spectrum of patients.^{54,55} In total, 18 studies were included in the final analysis. Studies were grouped according to

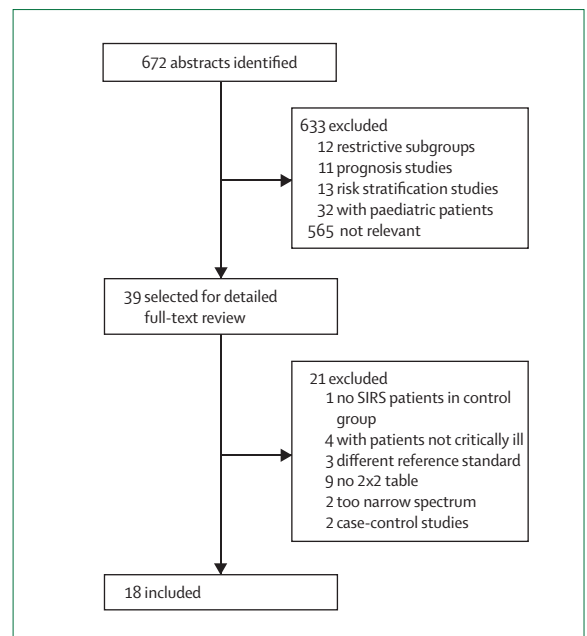


Figure 1: Study identification, inclusion, and exclusion

Some studies were excluded by more than one category. SIRS=systemic inflammatory response syndrome.

Sackett and Haynes' classification²⁹ for diagnostic studies: 14 phase 2 studies (group 1), four phase 3 studies (group 2). Details of all 18 studies are shown in table 2.

Study	Year	Country	Setting	Patients (n)	Mean age (years)	Cut-off point (ng/mL)	Study design	Prevalence of sepsis	Sensitivity	Specificity
Group 1 studies (1602 patients)										
Aikawa et al ⁵⁶	2005	Japan	Emergency department	176	47	0.5	PR	51%	0.64	0.86
Al-Nawas et al ¹⁶	1996	Germany	Hospital ward/ICU	337	..	0.5	PR	36%	0.60	0.79
Baumgarten et al ⁵⁷	2002	Netherlands	ICU	35	..	3	PR	31%	0.55	0.88
Chan et al ⁵⁸	2004	Taiwan	Emergency department	69	65	0.6	PR+CR	54%	0.71	0.67
Cheval et al ⁵⁹	2000	France	ICU	60	58	20	PR+CR	53%	0.88	0.82
Du et al ⁶⁰	2003	China	ICU	51	65	1.6	PR+CR	75%	0.80	0.74
Hausfater et al ⁶¹	2002	France	Emergency department	195	47	0.2	PR	35%	0.62	0.88
Muller et al ²³	2000	Switzerland	ICU	101	59	1.0	CR	58%	0.90	0.93
Mokart et al ⁶²	2005	France	ICU	50	56	1.1	PR	47%	0.81	0.74
Selberg et al ⁶³	2000	Germany	ICU	33	47	3.3	PR	67%	0.86	0.55
Suprin et al ⁶⁴	2000	France	ICU	95	57	2.00	PR	76%	0.65	0.70
Tugrul et al ⁶⁵	2002	Turkey	ICU	85	45	1.31	PR	88%	0.73	0.80
Ugarte et al ²¹	1999	Belgium	ICU	182	63	0.6	CR	58%	0.68	0.68
Wanner et al ⁶⁶	2000	Switzerland	ICU	133	40	1.5	PR	34%	0.76	0.77
Group 2 studies (495 patients)										
Bossink et al ⁶⁷	1999	Netherlands	Hospital ward/ICU	133	60	0.5	CR	45%	0.65	0.58
Gibot et al ⁶⁸	2004	France	ICU	76	60	0.6	PR+CR	62%	0.83	0.69
Harbarth et al ²²	2001	Switzerland	ICU	78	54	1.1	CR	77%	0.97	0.78
Ruokonen et al ²⁰	2002	Switzerland	ICU	208	55	0.8	PR+CR	78%	0.68	0.48

ICU=intensive care unit; PR=prospective recruitment; CR=consecutive recruitment; ..=not available.

Table 2: Characteristics of the studies included (2097 patients)

2097 patients were included in the analysis, with 1452 from intensive care units, 440 from emergency departments, and 205 from hospital wards. Studies included a wide case mix, including cardiac, pulmonary, neurological, gastrointestinal, renal, trauma, and surgical illnesses. SIRS criteria were fulfilled in 2092 patients. The mean age of patients in the studies was 54 years (range of study means 40–65 years). The prevalence of sepsis across studies ranged from 31% to 88%. All studies used LumiTest PCT, a commercially available immunoluminometric assay (Brahms Diagnostica, Berlin, Germany). Test threshold ranged from 0.2 ng/mL to 20 ng/mL.

Quantitative data synthesis

14 studies were included in group 1 (1602 patients). The pooled summary indices showed that the diagnostic performance of procalcitonin was low, with positive likelihood ratio 3.03 (95% CI 2.51–3.65), negative likelihood ratio 0.43 (95% CI 0.37–0.48), and diagnostic OR 7.79 (95% CI 5.86–10.35; figure 2). There was no evidence of a threshold effect ($b=0.451$, $p=0.66$). The SROC curve yielded a maximum joint sensitivity and specificity of 73% (95% CI 69–77), an area under the curve of 0.79, and Q^* point of 0.73, consistent with low diagnostic accuracy of procalcitonin.

One study had an unusually high summary estimate and accounted for most of the heterogeneity (52.6%).²³ Heterogeneity diminished significantly after this study was excluded (14.7%), thus allowing statistical pooling of the summary estimates. This study was therefore treated as an outlier and the results were reported with the exclusion of this study. However, subsequent sensitivity analysis showed that the pooled summary estimates did not differ significantly with inclusion of the outlier.

Four studies were included in group 2 (495 patients). These studies were highly heterogeneous (Cochran's $Q=21.57$, $p<0.001$), with an I^2 value of 86.1%. Statistical pooling was therefore not done for this group.

Finally, all 18 studies were pooled. There was no evidence of a threshold effect ($b=-0.21$, $p=0.40$). The SROC curve (figure 3) yielded a maximum joint sensitivity and specificity of 71% (95% CI 67–76), an area under the curve of 0.78, and Q^* point of 0.72, indicating that the performance of procalcitonin was low even when all studies were combined.

As expected, when pooling all studies, significant heterogeneity was introduced by the group 2 studies (Cochran's $Q=60.21$, $p<0.001$). The source of heterogeneity was explored by univariate meta-regression analysis. Sample size was significant in group 2 as a source of heterogeneity ($p=0.017$), but only weakly suggestive in group 1 ($p=0.09$; table 3). None of the variables, such as clinical settings, disease severity, patient demographics, or prevalence, were statistically significant as a source of variability in either group 1 or 2. Within group 2, smaller studies showed a higher diagnostic performance of procalcitonin (eg, a decrease

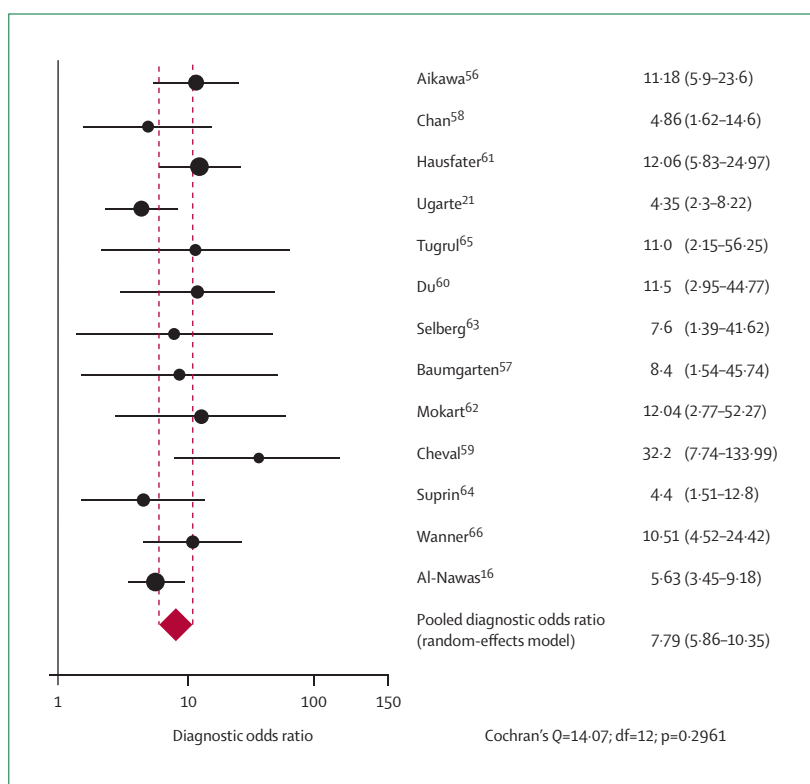


Figure 2: Diagnostic odds ratios of group 1 studies

Circles represent individual studies. Error bars represent 95% CIs. Diamond represents pooled diagnostic odds ratio, with dashed lines representing its 95% CI. Size of circles is proportional to weighting by inverse variance. SE=standard error.

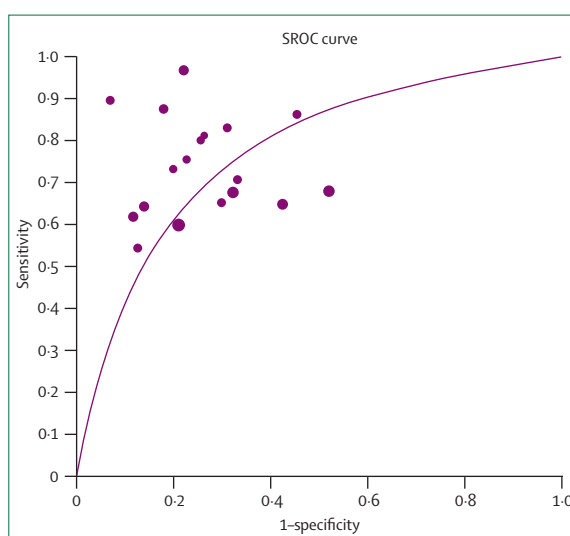


Figure 3: Summary receiver operator characteristic (SROC) curve of all studies

Circles indicate individual study estimates of sensitivity and 1-specificity. Size of circles is proportional to inverse variance of each study.

of 40 patients overestimated the relative diagnostic OR by a factor of 1.82). By contrast, the largest study (208 patients) had a diagnostic OR of 1.94 and a 95% CI that included the null effect of 1.0 (figure 4).

	Group 1		Group 2	
	Relative DOR (95% CI)	p	Relative DOR (95% CI)	p
Sample size*	1.00 (0.72–1.03)	0.092	0.55 (0.45–0.67)	0.017
Disease severity	0.93 (0.37–2.37)	0.480
Age	0.97 (0.91–1.03)	0.333	1.49 (0.73–3.08)	0.089
Study setting	1.04 (0.98–1.10)	0.217	0.90 (0.09–9.30)	0.660
Prevalence	1.00 (0.96–1.04)	0.926	0.94 (0.62–1.44)	0.321

..=not available. *The change in relative diagnostic odds ratio (DOR) is for an increase of 40 patients.

Table 3: Source of heterogeneity in univariate meta-regression analysis

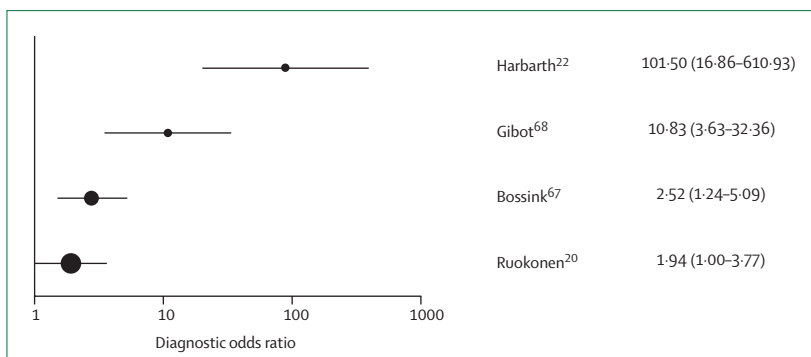


Figure 4: Diagnostic odds ratios of group 2 studies

Circles represent individual studies. Error bars represent 95% CIs. Size of circles is proportional to sample size.

Publication bias was detected using Egger's regression model ($p=0.006$). Visual inspection of the funnel plot suggested that missing studies were likely to fall to the left of the summary estimate. These studies were then imputed to calculate a new summary estimate (figure 5). The new diagnostic OR was 5.71 (95% CI 3.62–9.03), which was significantly lower than the observed diagnostic OR of 8.71 (95% CI 5.63–13.47). Therefore, the existing studies could have overestimated the diagnostic performance of procalcitonin.

Discussion

The results of this systematic review and meta-analysis indicate that the procalcitonin test cannot accurately distinguish sepsis from SIRS in critically ill adult patients. The study population in this review included a case mix typically seen in medical, surgical, or general intensive care units, emergency departments, and hospital wards. The findings of this review are therefore applicable to common clinical settings in which critically ill patients are managed.

The studies were grouped according to Sackett and Haynes' classification,²⁹ which assessed an index test on a continuum of diagnostic uncertainty. This continuum allows a stepwise, systematic progression in diagnostic evaluation from a training set (group 1), in which the index test was developed in an ideal situation, to a validation set (group 2), in which its performance was tested in a more realistic clinical context. Such

classification therefore allows clinicians to make a more informed decision when assessing the generalisability of studies.^{69–71}

Most patients (76%) were included in group 1 studies. The diagnostic OR and likelihood ratios were consistently low across most studies in this group. As a general rule, a diagnostic OR of greater than 100 indicates high accuracy, 25–100 indicates moderate accuracy, and less than 25 indicates an unhelpful test.^{72–74} The pooled diagnostic OR of 7.79 showed that the procalcitonin test was unlikely to be helpful in assisting clinical decision making in this group of patients. With a pretest probability of 40% in adult intensive-care-unit patients, use of the procalcitonin test would only raise the post-test probability to 66%. This is insufficient to influence treatment decision (eg, to start antibiotics). Conversely, with a negative likelihood ratio of 0.43, the application of a procalcitonin test would reduce the post-test probability to only 0.23, which is not quite enough to rule out an infection.

The remaining patients (24%) were included in group 2 studies. These studies were the most informative for clinical practice, as they were designed to resemble real-life situations by restricting to patients who were most likely to be encountered by clinicians. Group 2 summary estimates showed lower accuracy and more variability. Sample size gave rise to most of the variability, with smaller studies showing higher summary estimates. Other variables, such as patient age or clinical setting, were likely to have caused variation in the diagnostic performance of procalcitonin. However, the small number of studies ($n=4$) means that there is a lack of power in detecting these effects. Overall, these data suggest that smaller studies tend to overestimate the effect size, a finding that has been recognised in the diagnostic study literature.⁷⁵ A well-designed prospective

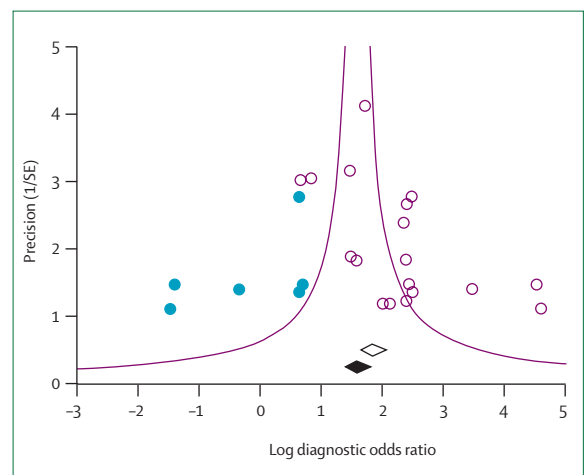


Figure 5: Publication bias detected by funnel plot

Unfilled circles indicate actual studies. Filled circles indicate imputed studies. Unfilled diamond indicates observed summary estimate. Black diamond indicates new summary estimate if all imputed studies were included. SE=standard error.

study with a larger sample size will be required to address this issue.

The diagnostic accuracy of procalcitonin in some populations of patients has recently been reviewed.^{76,77} Boysen and colleagues⁷⁶ assessed the diagnostic value of procalcitonin in post-operative infection. However, no conclusion could be drawn from their review because of significant heterogeneity among studies. Our analysis included one post-operative study,⁶² which was left out by this review. In another review, procalcitonin concentration was found to be better than C-reactive protein in diagnosing bacterial infection.⁷⁷ However, this review included studies across a wide range of age groups, clinical settings, and disease spectrum. Additionally, nearly half of the study population (46%) included paediatric patients and many patients did not have SIRS (57%). Despite such a diverse case mix, the study did not assess heterogeneity or its effect on the pooled estimates, thus making it very difficult to interpret its findings.⁷⁷ In view of these limitations, we applied in our study more strict inclusion criteria, focusing mainly on a more homogenous population, and used a substantially larger sample size (2092 vs 588). We also explored systematically the issue of heterogeneity by use of meta-regression and subgroup analysis. Furthermore, sensitivity analysis confirmed that our findings were robust and consistent. These methodological strengths have therefore enhanced the validity and applicability of our findings.

Publication bias is common in diagnostic studies and is possibly more of a problem than in studies of randomised controlled trials.⁷⁸ We detected publication bias in our review. As expected, the missing studies were located to the left of the funnel plot, consistent with the general observation that studies with less optimistic estimation of diagnostic performance are less likely to get published. With imputed values, the re-calculated diagnostic OR was significantly lower than the observed value, indicating that the true diagnostic performance of procalcitonin could have been even lower. However, the statistical methods used to assess publication bias have limitations.^{79,80} The above findings therefore need to be interpreted in this context.

The scope of this review means that our findings cannot be generalised to specific diseases (eg, pancreatitis, burns) or settings (eg, cardiothoracic surgical patients, neonatal/paediatric patients). Our study did not include patients who were not critically ill, or who did not fulfil the SIRS criteria. The variation in disease prevalence and severity in these patients means that the diagnostic accuracy of procalcitonin is likely to be different, depending on the chosen population or setting. Finally, we did not include studies that assessed the ability of procalcitonin to diagnose septic shock, since these conditions were usually recognised by simple clinical criteria.

The focus of this review is on the role of procalcitonin in distinguishing sepsis from SIRS in critically ill patients.

However, infection can be present without any clinical manifestation of SIRS.⁸¹ The role of procalcitonin in such a setting remains undefined, since most of the procalcitonin studies in this review used SIRS patients in the control groups. Furthermore, this review does not address the issue of prognosis. Further studies would be needed to assess the role of procalcitonin in both these settings.

Although the SIRS criteria are widely used in the literature surveyed by this review, they have been criticised for being too sensitive.⁸² However, this low threshold for detection is appropriate for a test for which the consequences of overdetection are outweighed by the consequences of undetection for potentially septic patients.⁸³ Additionally, the SIRS criteria provide uniformity in inclusion criteria and allow valid comparison to be made across many different studies.⁸⁴ Such uniformity has ensured the validity of statistical pooling in our meta-analysis. Despite its limitations, the continuing use of the SIRS concept has recently been supported by an international sepsis definitions conference.¹⁷ The findings of our study therefore reflect the prevalent use of the SIRS concept in sepsis research.^{85,86}

Ideally, the additive value of the procalcitonin test to supplement a clinician's bedside assessment should be evaluated in any diagnostic study. Unfortunately, most of the 18 studies did not explore how procalcitonin could be used to enhance clinical assessment, which highlights a recent trend of adopting a biomarker-based approach to diagnose sepsis. In light of our findings, future research should focus on incorporating biomarkers as part of an overall assessment of critically ill patients, rather than in preference to clinical assessment.

In summary, we found that procalcitonin had a low diagnostic performance in differentiating sepsis from SIRS in critically ill adult patients. The evidence presented in this review does not lend support to the widespread use of the procalcitonin test for sepsis diagnosis in critical care settings.

Conflicts of interest

We declare that we have no conflicts of interest.

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References

- 1 Martin G, Mannino D, Eaton S, Moss M. The epidemiology of sepsis in the United States from 1979 through 2000. *N Engl J Med* 2003; **348**: 1546–54.
- 2 Warren H. Strategies for the treatment of sepsis. *N Engl J Med* 1997; **336**: 952–53.
- 3 Parillo J, Parker M, Natanson C, et al. Septic shock: advances in the understanding of pathogenesis, cardiovascular dysfunctions, and therapy. *Ann Intern Med* 1990; **113**: 227–42.
- 4 Pittet D, Rangel-Frausto S, Li N, Tarara D, Costigan M. Systemic inflammatory response syndrome, sepsis, severe sepsis and septic shock: incidence, morbidities and outcomes in surgical ICU patients. *Intensive Care Med* 1995; **21**: 302–09.

Search strategy and selection criteria

These are described in detail in the Methods section on page 210.

- 5 Kollef M, Sherman G, Ward S, Fraser V. Inadequate antimicrobial treatment of infections: a risk factor for hospital mortality among critically ill patients. *Chest* 1999; **115**: 462–74.
- 6 Bochud P-Y, Glauser MP, Calandra T. Antibiotics in sepsis. *Intensive Care Med* 2001; **27** (suppl 1): S33–48.
- 7 Brun-Buisson C. The epidemiology of the systemic inflammatory response. *Intensive Care Med* 2000; **26** (suppl 1): S64–74.
- 8 American College of Chest Physicians, Society of Critical Care Medicine Consensus Conference. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Crit Care Med* 1992; **20**: 864–74.
- 9 Rangel-Frausto M, Pittet D, Costigan M, Hwang T, Davis C, Wenzel R. The natural history of the systemic inflammatory response syndrome (SIRS). A prospective study. *JAMA* 1995; **273**: 117–23.
- 10 Marshall J. Infection and the host septic response: implication for clinical trials of mediator antagonism. In: Vincent J-L, ed. *Yearbook of intensive care and emergency medicine*. Berlin: Springer-Verlag, 1994: 3–13.
- 11 Brun-Buisson C, Doyon F, Carlet J, et al. Incidence, risk factors, and outcome of severe sepsis and septic shock in adults. A multicenter prospective study in intensive care units. *JAMA* 1995; **274**: 968–74.
- 12 Young L. Gram-negative sepsis. In: Mandell G, Douglas R, Bennet J, eds. *Principles and practice of infectious diseases*. New York: Churchill Livingstone, 1990: 611–36.
- 13 Assicot M, Gendrel D, Carsin H, et al. High serum procalcitonin concentration in patients with sepsis and infection. *Lancet* 1993; **341**: 515–18.
- 14 Brunkhorst FM, Wegscheider K, Forycki ZF, Brunkhorst R. Procalcitonin for early diagnosis and differentiation of SIRS, sepsis, severe sepsis, and septic shock. *Intensive Care Med* 2000; **26** (suppl 2): S148–52.
- 15 Aouifi A, Piriou V, Bastien O, et al. Usefulness of procalcitonin for diagnosis of infection in cardiac surgical patients. *Crit Care Med* 2000; **28**: 3171–76.
- 16 Al-Nawas B, Krammer I, Shah PM. Procalcitonin in diagnosis of severe infections. *Eur J Med Res* 1996; **1**: 331–33.
- 17 Levy M, Fink M, Marshall J, et al. 2001 SCCM/ESICM/ACCP/ATS/SIS International Sepsis Definitions Conference. *Crit Care Med* 2003; **31**: 1250–56.
- 18 Vincent J-L. THE marker of sepsis? *Crit Care Med* 2000; **28**: 1226–28.
- 19 Lapillonne A, Basson E, Monneret G, Biennu J, Salle B. Lack of specificity of procalcitonin for sepsis diagnosis in premature infant. *Lancet* 1998; **351**: 1211–12.
- 20 Ruokonen E, Ilkka L, Niskanen M, Takala J. Procalcitonin and neopterin as indicators of infection in critically ill patients. *Acta Anaesthesiol Scand* 2002; **46**: 398–404.
- 21 Ugarte H, Silva E, Mercan D, De Mendonca A, Vincent J-L. Procalcitonin used as a marker of infection in the intensive care unit. *Crit Care Med* 1999; **27**: 498–504.
- 22 Harbarth S, Holeckova K, Froidevaux C, et al. Diagnostic value of procalcitonin, interleukin-6, and interleukin-8 in critically ill patients admitted with suspected sepsis. *Am J Respir Crit Care Med* 2001; **164**: 396–402.
- 23 Muller B, Becker KL, Schachinger H, et al. Calcitonin precursors are reliable markers of sepsis in a medical intensive care unit. *Crit Care Med* 2000; **28**: 977–83.
- 24 Christ-Crain M, Jaccard-Stolz D, Bingisser R, et al. Effect of procalcitonin-guided treatment on antibiotic use and outcome in lower respiratory tract infections: cluster-randomised, single-blinded intervention trial. *Lancet* 2004; **363**: 600–07.
- 25 Deville W, Buntinx F. Guidelines for conducting systematic reviews of studies evaluating the accuracy of diagnostic tests. In: Knottnerus J, ed. *The evidence base of clinical diagnosis*, 1st edn. London: BMJ, 2002: 145–65.
- 26 Lijmer J, Mol B, Heisterkamp S, et al. Empirical evidence of design-related bias in studies of diagnostic tests. *JAMA* 1999; **282**: 1061–66.
- 27 Whiting P, Rutjes A, Reitsma J, Bossuyt P, Kleijnen J. The development of QUADAS: a tool for the quality assessment of studies of diagnostic accuracy included in systematic reviews. *BMC Med Res Methodol* 2003; **3**: 1–13.
- 28 Whiting P, Rutjes A, Reitsma J, Bossuyt P, Kleijnen J. Development and validation of methods for assessing the quality of diagnostic accuracy studies. *Health Technol Assess* 2004; **8**: 1–264.
- 29 Sackett D, Haynes R. The architecture of diagnostic research. In: Knottnerus J, ed. *The evidence base of clinical diagnosis*, 1st edn. London: BMJ, 2002: 19–38.
- 30 Glas A, Lijmer J, Prins M, Bonsel G, Bossuyt P. The diagnostic odds ratio: a single indicator of test performance. *J Clin Epidemiol* 2003; **56**: 1129–35.
- 31 DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials* 1986; **7**: 177–88.
- 32 Moses L, Littenberg B, Shapiro D. Combining independent studies of a diagnostic test into a summary ROC curve: data-analytical approaches and some additional consideration. *Stat Med* 1993; **12**: 1293–316.
- 33 Walter S. Properties of the summary receiver operating characteristics (SROC) curve for diagnostic test data. *Stat Med* 2002; **21**: 1237–56.
- 34 Egger M, Davey S, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* 1997; **315**: 629–34.
- 35 Duval S, Tweedie R. Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. *Biometrics* 2000; **56**: 455–63.
- 36 Giamarellos-Bourboulis EJ, Mega A, Grecka P, et al. Procalcitonin: a marker to clearly differentiate systemic inflammatory response syndrome and sepsis in the critically ill patient? *Intensive Care Med* 2002; **28**: 1351–56.
- 37 Chirouze C, Schuhmacher H, Rabaud C, et al. Low serum procalcitonin level accurately predicts the absence of bacteremia in adult patients with acute fever. *Clin Infect Dis* 2002; **35**: 156–61.
- 38 Delevaux I, Andre M, Colombier M, et al. Can procalcitonin measurement help in differentiating between bacterial infection and other kinds of inflammatory processes? *Ann Rheum Dis* 2003; **62**: 337–40.
- 39 Munoz P, Simarro N, Rivera M, Alonso R, Alcalá L, Bouza E. Evaluation of procalcitonin as a marker of infection in a nonselected sample of febrile hospitalized patients. *Diagn Microbiol Infect Dis* 2004; **49**: 237–41.
- 40 Stucker F, Herrmann F, Graf J-D, Michel J, Krause K, Gauazzi G. Procalcitonin and infection in elderly patients. *J Am Geriatr Soc* 2005; **53**: 1392–95.
- 41 Geppert A, Steiner A, Delle-Karth G, Heinz G, Huber K. Usefulness of procalcitonin for diagnosing complicating sepsis in patients with cardiogenic shock. *Intensive Care Med* 2003; **29**: 1384–89.
- 42 Liaudat S, Dayer E, Praz G, Bille J, Troillet N. Usefulness of procalcitonin serum level for the diagnosis of bacteremia. *Eur J Clin Microbiol Infect Dis* 2001; **20**: 524–27.
- 43 Aalto H, Takala A, Kautiainen H, Repo H. Laboratory markers of systemic inflammation as predictors of bloodstream infection in acutely ill patients admitted to hospital in medical emergency. *Eur J Clin Microbiol Infect Dis* 2004; **23**: 699–704.
- 44 Caterino J, Scheatzle M, Forbes M, D'Antonio J. Bacteremic elder emergency department patients: procalcitonin and white count. *Acad Emerg Med* 2004; **11**: 393–96.
- 45 Guven H, Altintop L, Baydin A, et al. Diagnostic value of procalcitonin levels as an early indicator of sepsis. *Am J Emerg Med* 2002; **20**: 203–06.
- 46 Bell K, Wattie M, Byth K, et al. Procalcitonin: a marker of bacteraemia in SIRS. *Anaesth Intensive Care* 2003; **31**: 629–36.
- 47 Brunkhorst FM, Eberhard OK, Brunkhorst R. Discrimination of infectious and noninfectious causes of early acute respiratory distress syndrome by procalcitonin. *Crit Care Med* 1999; **27**: 2172–76.
- 48 Luzzani A, Polati E, Dorizzi R, Rungtatscher A, Pavan R, Merlini A. Comparison of procalcitonin and C-reactive protein as markers of sepsis. *Crit Care Med* 2003; **31**: 1737–41.
- 49 Balci C, Sungurtekin H, Gurses E, Sungurtekin U, Kaptanoglu B. Usefulness of procalcitonin for diagnosis of sepsis in the intensive care unit. *Crit Care Med* 2003; **7**: 85–90.
- 50 Prucha M, Herold I, Zazula R, Dubska L, Kavka B, Dosta M. A comparison of procalcitonin, interleukin-6 and C-reactive protein in differential diagnosis of patients with the septic syndrome treated at the intensive care units. *Vnitř Lek* 2003; **49**: 541–47 (in Czech).
- 51 Kazda A, Valenta J, Brodská H, Stach Z, Hendl J, Stritesky M. Procalcitonin in critically ill patients. *Klin Biochem Metabol* 2005; **13**: 4–9 (in Czech).
- 52 Hergert M, Lestin H-G, Scherkus M, et al. Procalcitonin in patients with sepsis and polytrauma. *Clin Lab* 1998; **44**: 659–70.

- 53 Castelli GP, Pognani C, Meisner M, Stuani A, Bellomi D, Sgarbi L. Procalcitonin and C-reactive protein during systemic inflammatory response syndrome, sepsis and organ dysfunction. *Crit Care* 2004; **8**: R234–42.
- 54 Clec'h C, Ferriere F, Karoubi P, et al. Diagnostic and prognostic value of procalcitonin in patients with septic shock. *Crit Care Med* 2004; **32**: 1166–69.
- 55 Reith HB, Mittelkotter U, Wagner R, Thiede A. Procalcitonin (PCT) in patients with abdominal sepsis. *Intensive Care Med* 2000; **26**: S165–69.
- 56 Aikawa N, Fujishima S, Endo S, et al. Multicenter prospective study of procalcitonin as an indicator of sepsis. *J Infect Chemother* 2005; **11**: 152–59.
- 57 Baumgarten R, Pequeriaux NCV, Van Puyenbroek MJE, Speelberg B. Diagnosis of sepsis by procalcitonin. *Ned Tijdschr Klin Chem* 2002; **27**: 32–35.
- 58 Chan YL, Tseng CP, Tsay PK, Chang SS, Chiu TF, Chen JC. Procalcitonin as a marker of bacterial infection in the emergency department: an observational study. *Crit Care* 2004; **8**: R12–20.
- 59 Cheval C, Timsit JF, Garrouste-Orgeas M, et al. Procalcitonin (PCT) is useful in predicting the bacterial origin of an acute circulatory failure in critically ill patients. *Intensive Care Med* 2000; **26** (suppl 2): S153–58.
- 60 Du B, Pan J, Cheng D, Li Y. Serum procalcitonin and interleukin-6 levels may help to differentiate systemic inflammatory response of infectious and non-infectious origin. *Chin Med J* 2003; **116**: 538–42.
- 61 Hausfater P, Garric S, Ayed B, Rosenheim M, Bernard M, Riou B. Usefulness of procalcitonin as a marker of systemic infection in emergency department patients: a prospective study. *Clin Infect Dis* 2002; **34**: 895–901.
- 62 Mokart D, Merlin M, Sannini A, et al. Procalcitonin, interleukin 6 and systemic inflammatory response syndrome (SIRS): early markers of postoperative sepsis after major surgery. *Br J Anaesth* 2005; **94**: 767–73.
- 63 Selberg O, Hecker H, Martin M, Klos A, Bautsch W, Kohl J. Discrimination of sepsis and systemic inflammatory response syndrome by determination of circulating plasma concentrations of procalcitonin, protein complement 3a, and interleukin-6. *Crit Care Med* 2000; **28**: 2793–98.
- 64 Suprin E, Camus C, Gacouin A, et al. Procalcitonin: a valuable indicator of infection in a medical ICU? *Intensive Care Med* 2000; **26**: 1232–38.
- 65 Tugrul S, Esen F, Celebi S, et al. Reliability of procalcitonin as a severity marker in critically ill patients with inflammatory response. *Anaesth Intensive Care* 2002; **30**: 747–54.
- 66 Wanner GA, Keel M, Steckholzer U, Beier W, Stocker R, Ertel W. Relationship between procalcitonin plasma levels and severity of injury, sepsis, organ failure, and mortality in injured patients. *Crit Care Med* 2000; **28**: 950–57.
- 67 Bossink A, Groeneveld A, Thijs LG. Prediction of microbial infection and mortality in medical patients with fever: plasma procalcitonin, neutrophilic elastase-alpha-antitrypsin, and lactoferrin compared with clinical variables. *Clin Infect Dis* 1999; **29**: 398–407.
- 68 Gibot S, Kolopp-Sarda M-N, Bene M-C. Plasma level of a triggering receptor expressed on myeloid cells-1: its diagnostic accuracy in patients with suspected sepsis. *Ann Intern Med* 2004; **141**: 9–15.
- 69 Tatsioni A, Zarin D, Aronson N, Samson D, et al. Challenges in systematic reviews of diagnostic technologies. *Ann Intern Med* 2005; **142**: 1048–55.
- 70 Nierenberg A, Feinstein A. How to evaluate a diagnostic marker test. Lessons from the rise and fall of dexamethasone suppression test. *JAMA* 1988; **259**: 1699–702.
- 71 Jenicek M. Identifying cases of disease. Clinimetrics and diagnosis. In: Jenicek M, ed. Foundations of evidence-based medicine. London: Parthenon, 2003: 107–46.
- 72 Jaeschke R, Guyatt G, Sackett D. User's guides to the medical literature. III. How to use an article about a diagnostic test. B. What are the results and will they help me in caring for my patients? The Evidence-Based Medicine Working Group. *JAMA* 1994; **271**: 703–07.
- 73 Deeks J. Systematic reviews of evaluations of diagnostic and screening tests. In: Egger M, Smith GD, Altman DG, eds. Systematic reviews in health care: meta-analysis in context, 2nd edn. London: BMJ, 2001: 248–82.
- 74 Swets J. Measuring the accuracy of diagnostic systems. *Science* 1988; **240**: 1285–93.
- 75 Song F, Khan KS, Dinnes J, Sutton AJ. Asymmetric funnel plots and publication bias in meta-analyses of diagnostic accuracy. *Int J Epidemiol* 2002; **31**: 88–95.
- 76 Boysen A, Madsen JS, Jorgensen PE. Procalcitonin as a marker of postoperative complications. *Scand J Clin Lab Invest* 2005; **65**: 387–94.
- 77 Simon L, Gauvin F, Amre D, Saint-Louis P, Lacroix J. Serum procalcitonin and C-reactive protein levels as markers of bacterial infection: a systematic review and meta-analysis. *Clin Infect Dis* 2004; **39**: 206–17.
- 78 Irwig L, Macaskill P, Glasziou P, Fahey M. Meta-analytical methods for diagnostic test accuracy. *J Clin Epidemiol* 1995; **48**: 119–30.
- 79 Sterne J, Gavaghan D, Egger M. Publication and related biases in meta-analysis: power of statistical tests and prevalence in the literature. *J Clin Epidemiol* 2000; **53**: 1119–29.
- 80 Irwig L, Berry G. Graphical test is itself biased. *BMJ* 1998; **316**: 469–69.
- 81 Alberti C, Brun-Buisson C, Goodman SV, et al. Influence of systemic inflammatory response syndrome and sepsis on outcome of critically ill infected patients. *Am J Respir Crit Care Med* 2003; **168**: 77–84.
- 82 Abraham E, Matthay M, Dinarello C, et al. Consensus conference definitions for sepsis, septic shock, acute lung injury, and acute respiratory distress syndrome: time for a reevaluation. *Crit Care Med* 2000; **28**: 232–35.
- 83 Bone R, Balk R, Cerra F, et al. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. The ACCP/SCCM Consensus Conference Committee. American College of Chest Physicians/Society of Critical Care Medicine. *Chest* 1992; **101**: 1644–55.
- 84 Sands K, Bates D, Lanken P, Graman P, et al. Epidemiology of sepsis syndrome in 8 academic medical centers. *JAMA* 1997; **277**: 234–40.
- 85 Trzeciak S, Zanotti-Cavazzoni S, Parillo J, Dellinger R. Inclusion criteria for clinical trials in sepsis: did the American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference Definitions of Sepsis have an impact? *Chest* 2005; **127**: 242–45.
- 86 Sprung C, Sakr Y, Vincent J-L, et al. An evaluation of systemic inflammatory response syndrome signs in the Sepsis Occurrence in Acutely ill Patients (SOAP) study. *Intensive Care Med* 2006; **32**: 421–27.